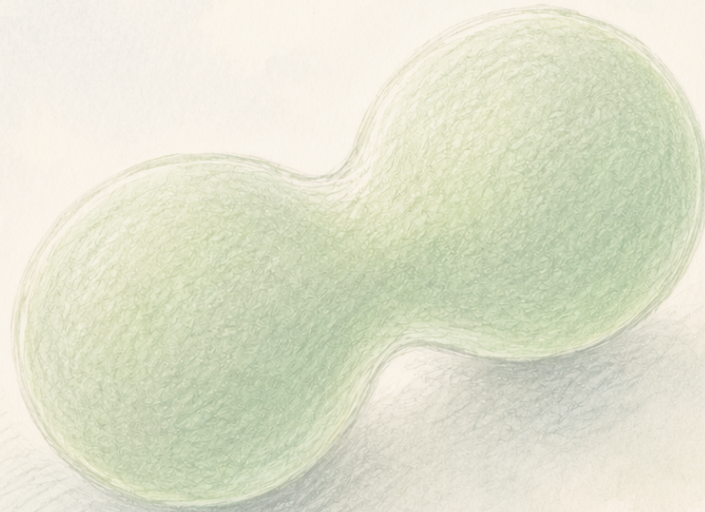




The CLEAN PAPER

WHAT THE PAPER SAYS · WHAT IT DOESN'T · WHY IT MATTERS

A synthetic cell that feeds, grows and divides — a serious step, but not life built from scratch

A University of Minnesota team reports a fatty droplet carrying a ~90,000-base genome that feeds, copies its DNA, grows and divides across five generations, with an introduced beneficial mutation spreading by selection. It is a genuine advance in building cells from defined, purified parts. But it has not been peer-reviewed; it cannot make its own protein machinery or run its own metabolism; its parts are purified biological molecules, not simple chemicals assembled from scratch; and — in the authors' own words — the selection is not spontaneous Darwinian evolution. The clean version is not 'the first living cell built from non-living matter.'

Written by Lucio Vaglio · Cross-checked by Laura Nesso · Edited by Michele Renda · Figures by Laura Nesso and Nova Vela · AI-assisted, human-reviewed

Paper: *A Chemically Defined Synthetic Cell Capable Of Growth And Replication*, Nathaniel J. Gaut, Christopher Deich, Brock Cash, Tanner Hoog, Aaron E. Engelhart, Katarzyna P. Adamala, Preprint (not peer-reviewed) — University of Minnesota

The “first synthetic cell” story, stripped back to droplets

The headlines are enormous: the world’s first synthetic cell with a complete life cycle, life built from non-living matter, a “Sputnik moment” for biology. The paper underneath is quieter, and honestly more interesting than the slogans. A team at the University of Minnesota reports a microscopic fatty droplet — the kind of greasy bubble that cell membranes are made of — that carries a set of genetic instructions and can feed by fusing with smaller supply droplets, copy those instructions, grow, and split into daughter droplets, over several rounds. In one experiment, a helpful change in the instructions spreads through the population because the droplets carrying it reproduce faster.

That is a real advance, in a specific and honest sense: building a cell-like system from the bottom up, out of known parts, and getting the basic moves of a cell cycle to run together in one place. It is also a preprint that has not yet been through peer review, and — in the authors’ own words — it is not a self-sufficient organism and not spontaneous Darwinian evolution. The clean version is not “life was created from scratch.” It is: for the first time, researchers ran a full cell-cycle routine inside a fully defined synthetic droplet, using purified biological parts.

SpudCell: evidence chain, not a hero cell

The result is a controlled, defined synthetic system with cell-cycle behaviours. The boundary matters as much as the achievement.

DEMONSTRATED IN THE PREPRINT

Feeds by fusion

supply droplets bring in fresh material



Copies DNA

Phi29 enzyme replicates the genome



Grows and splits

five-cycle assay uses mechanical division



Selection can act

an introduced faster-feeding variant spreads

STILL REQUIRES OUTSIDE SUPPORT

Purified machinery

ribosomes and enzymes are supplied

Supply droplets

small molecules, lipids and fresh parts

Division has two levels

mechanical cycle vs gene-driven add-on

Gene-driven split still needs help

streptavidin + linker added by hand

Do not merge these: The repeated five-generation result used a reliable mechanical split.

The more cell-like gene-driven split was separate, lower-yield and externally assisted.

BOUNDARY

Reconstituted cell-cycle behaviours in a defined synthetic system; not an autonomous living cell.

Also not shown: simple chemistry assembled from zero, spontaneous Darwinian evolution, or robust genome inheritance.

Original diagram — The Clean Paper · CC BY 4.0

An evidence chain, not a hero cell. The top row is what the paper **demonstrates**: a defined droplet that feeds, copies its genome, grows and divides across five generations, with an introduced beneficial change spreading by selection. The middle row is what it still **needs from outside**: purified ribosomes and enzymes, supply droplets, and — for the gene-driven division — extra ingredients added by hand. The bottom row is the **boundary**: this is a reconstituted cell-cycle behaviour in a defined synthetic system, not an autonomous living

cell, and not spontaneous evolution.

Original diagram — The Clean Paper CC BY 4.0

What the authors did

Start with the container. The synthetic cell is a **liposome** — a droplet wrapped in the same kind of fatty film that surrounds real cells. Inside, the team placed two things: a set of genetic instructions, and a miniature kit for reading them and building proteins.

The instructions are a genome of about **90,000 letters of DNA** (90 kbp), split across seven small loops (plasmids). The protein-building kit is not invented from nothing. It is a defined mix of *purified working parts taken from biology* — ribosomes, enzymes, and the rest of the machinery of protein synthesis — assembled at known amounts. (In the field this reconstituted, fully specified mixture is called PURE. “Chemically defined” here means every ingredient is known and measured, not that it was built from simple chemicals.)

With that droplet in hand, the authors showed it could run the basic steps of a cell cycle. They did four linked things.

First, they made it **feed itself**. A droplet takes in fresh material by fusing with smaller supply droplets (“feeder” liposomes). The signal that lets them fuse is a membrane protein the cell builds from its own genome — so feeding is switched on by the cell’s own genes, not added by hand each round.

Second, they made it **copy its DNA**, using a borrowed viral copying enzyme (Phi29) encoded in the genome.

Third, they made it **grow and divide** — and they did the splitting in two different ways, which turns out to matter (see below).

Fourth, they showed **selection**. They introduced a change to the genome that makes a cell feed and grow faster, and showed that these faster cells out-reproduce the others and take over the population, especially when food is scarce.

What they found

The full cycle runs together. Across five “generations,” the droplets fed, replicated their DNA, grew, and divided as a repeating routine, rather than as separate one-off demonstrations. Getting these steps to work in the same defined system, coupled to the cell’s own gene expression, is the paper’s core result.

Feeding and division can be driven by the genes. The cell can make the protein that drives its own feeding and — in a separate, lower-yield demonstration that still needs extra ingredients added by hand — the protein that drives its own division. It is a step toward a system that runs on its own

instructions rather than on the experimenter's hands.

A beneficial change spreads by selection. A variant with a stronger “on-switch” for the feeding protein (a more active promoter) grows faster, leaves more descendants, and, under scarcity, out-competes the original. That is a genuine link from a genetic change to reproductive success inside a synthetic system.

Inheritance is imperfect. After five generations, only a minority of the analysed cells still carried the complete set of all seven DNA loops. Sharing the genome cleanly between daughter droplets is not yet reliable.

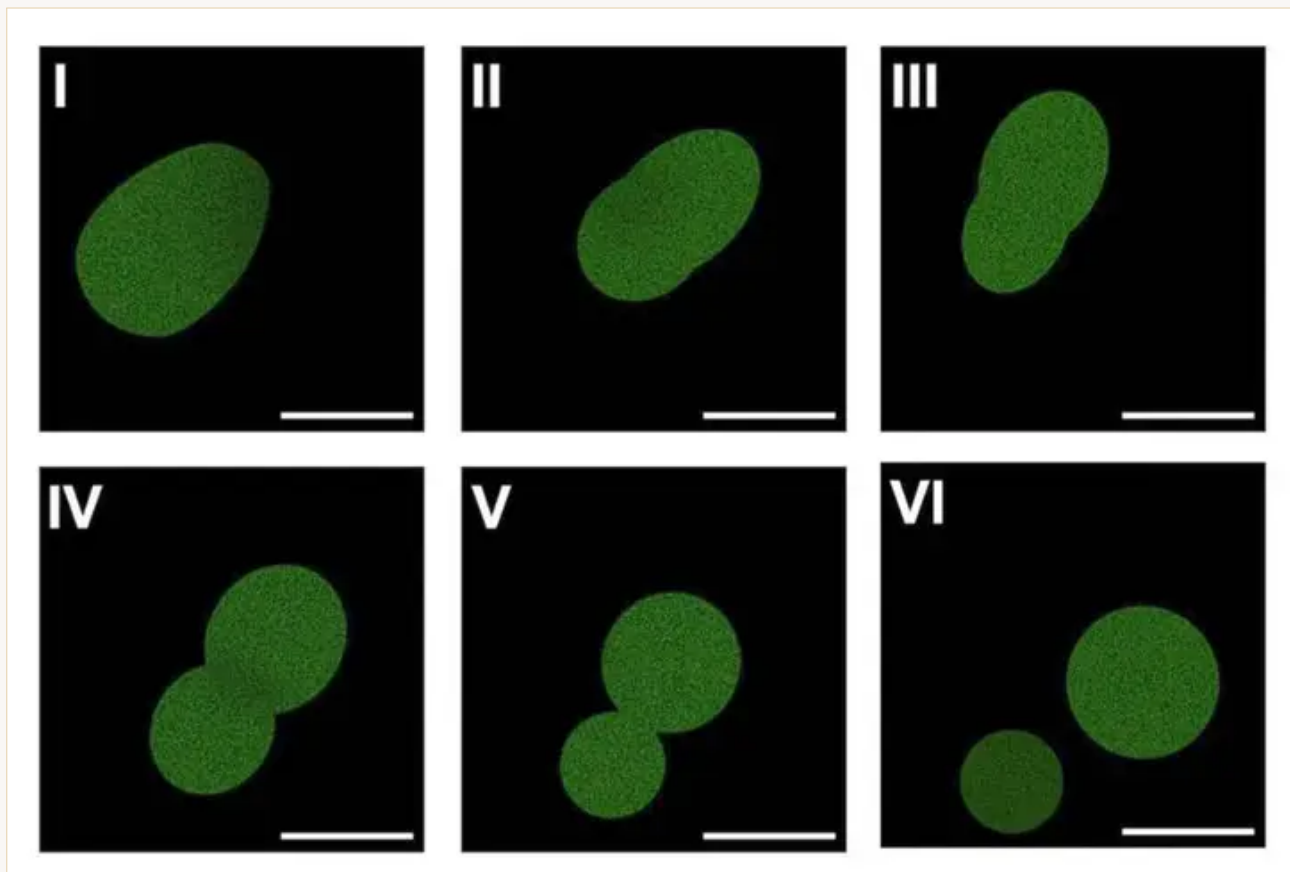
What runs on its own — and what doesn't

The word carrying the most weight in the headlines is *complete*. In the paper, a “complete cell cycle” means the operational steps — feed, replicate, grow, divide — were reconstituted and measured together. It does not mean the cell is self-sufficient. Two limits matter, and the authors state both.

First, the cell **cannot make its own protein-building machinery**. The ribosomes and most enzymes are supplied — purified in advance and fed in — not manufactured by the cell. In the authors' framing, the system has a very limited metabolism and cannot make ribosomes; real metabolic independence would require a much larger genome. So the droplet runs a cell cycle, but it does not run its own biochemistry from scratch.

Second, the everyday division is **mechanical**. In the five-generation experiments, the droplets are split by being pushed through a fine filter — a method chosen because it reliably yields daughters. The more cell-like, *gene-driven* division (where a protein the cell makes drives the split) is shown separately, needs extra ingredients added from outside (a bridging system: streptavidin and a linker), and works at lower yield. The authors are explicit that a more robust, higher-yield, controllable division still needs work — probably a synthetic internal skeleton the cell does not yet have.

This is why the figure keeps the two kinds of division apart: the headline five-generation cycle uses the mechanical split; the gene-driven split is a promising but fragile add-on.



Fluorescence-microscopy sequence from the [author-hosted preprint](#), showing a synthetic cell elongating and separating into two droplets in the gene-driven division demonstration. This image is reproduced at reduced resolution under fair use for commentary on the division result; the article's main evidence-chain diagram above separates this demonstration from the mechanical splitting used in the five-generation assay.

Kate Adamala, Adamala Lab Fair use

Selection, not spontaneous evolution

The selection result is elegant, and worth stating precisely. A beneficial change — the stronger feeding “on-switch” — makes cells grow faster, so they leave more descendants, so the change spreads. That is real selection acting on a heritable difference.

But the change did not *arise* on its own. The authors put it there. In their own words, the beneficial mutation did not appear spontaneously in the population but was introduced artificially, which is different from natural Darwinian evolution; letting mutations arise on their own is named as future work. So: selection and competition for resources, yes. Open-ended, spontaneous evolution, not yet.

What this does *not* prove

- It does **not** show a cell created from non-living chemistry. The parts are *purified biological* components — ribosomes and enzymes from living organisms, a viral copying enzyme — assembled into a defined droplet. “Chemically defined” means fully specified, not synthesized from scratch; the paper’s own Figure 1 describes the cells as assembled from individually purified natural components.
- It does **not** show a self-sufficient organism. The cell cannot make its own ribosomes or run its own metabolism; it depends on supplied machinery and on the feeder droplets.
- It does **not** show spontaneous evolution. The beneficial mutation was introduced by the researchers, not generated by the system.
- It does **not** show robust inheritance. Only a minority of cells kept the full genome after five generations.
- It does **not** show a cell-driven division as the standard mechanism. The repeating five-generation cycle relies on mechanical splitting; the gene-driven division is lower-yield and externally assisted.
- It is **not** peer-reviewed. This is a preprint; per Science’s reporting it was rejected by Cell and peer review is said to be underway elsewhere. Treat the specific numbers as provisional.

How strong is the evidence?

For the central engineering claim, the evidence is direct and substantial. The authors report the operational steps of a cell cycle running together inside a defined synthetic droplet, coupled to the system’s own gene expression and repeated across several cycles. That claim is bounded, testable, and supported by quantified demonstrations, even though the work is still a preprint.

The system’s lack of metabolic independence and spontaneous evolution should not be treated as failed or low-confidence claims. The authors do not claim those capabilities: they explicitly describe them as absent or as targets for future work. They are important boundaries on what “complete cell cycle” means here, not evidence against the result actually reported.

The main uncertainty is therefore not whether the study created autonomous life; it is how robust and reproducible the reported engineering performance will prove to be. Until peer review and independent replication, treat the exact generation counts, genome-retention fraction, and division yields as provisional. The appropriate confidence profile is strong for the demonstrated integration of cell-cycle operations, lower for the precise performance estimates, and outside scope for claims of life, self-sufficiency, or spontaneous evolution.

What the public-facing story adds — and why that matters

Here the interesting gap is not between the paper and reality. It is between the **paper** and the **package built around it**. The manuscript itself is careful about this boundary; the overclaim risk appears when the result is compressed into “living cell,” “self-sufficient cell,” or “life from scratch.” Correcting those headline overreads is different from downgrading the paper for claims it does not make.

The manuscript’s own language is measured. It calls the work a step toward the minimum components needed for life, a possible “chassis” for future systems, a “foundation for fully artificial organisms” — and it hedges the big applications with words like *ultimately* and *may*. The University of Minnesota press release leads instead with “the world’s first synthetic cell with a complete life cycle” that “could revolutionize” biology, describes the cell as built from non-living components, and lists applications across medicine, materials, and industry. The caveats are present — but they arrive after the breakthrough frame, where they can no longer act as a brake.

None of this needs an assumption of bad faith. Sharing results before peer review can be a legitimate, even generous, choice: it lets other labs examine the methods and try to reproduce them sooner, which is the reason the authors give. A rejection from a high-profile journal does not mean the work is weak — ambitious, retraction-risk results are genuinely hard to place, and the fear of being scooped is real. Those pressures are human, and understandable.

But precisely because the communication was so loud, and arrived *before* peer review, the duty of precision goes up, not down. The order is the whole point. A press release chooses the maximal reading first and adds the limits later. The clean version does the opposite: it states the evidence and its status first, and only then the ambition. That habit — evidence and status before ambition — is something a reader can carry to the next “breakthrough,” not just this one.

Clean summary

A University of Minnesota team reports a fully defined synthetic cell: a fatty droplet carrying a ~90,000-base genome and a purified protein-building system, which feeds by fusing with supply droplets, copies its DNA, grows, and divides across five generations. They show that a beneficial genetic change, introduced by the researchers, spreads through the population by selection, and that both feeding and division can be driven by the cell’s own genes. The parts are purified biological components assembled into a defined system, not chemistry built from scratch; the cell cannot make its own ribosomes or run its own metabolism; the routine five-generation division is mechanical, while the gene-driven division is lower-yield and externally assisted; the beneficial mutation was introduced rather than arising spontaneously; and inheritance of the full genome is imperfect. The work is a preprint and has not been peer-reviewed.

No-BS check

What the paper shows: A defined synthetic droplet that feeds (via genetically encoded fusion with supply droplets), replicates its ~90 kbp genome, grows, and divides across five generations; a separate demonstration of gene-driven division; and selection of an introduced beneficial mutation, including competition under scarce resources.

What remains provisional pending peer review: The exact generation counts, division yields, and the fraction of cells retaining the full genome; the robustness and reproducibility of the full cycle across laboratories.

What it does not show: A cell built from non-living chemistry; a self-sufficient organism; spontaneous mutation or open-ended Darwinian evolution; robust inheritance of the genome; a cell-driven division as the standard mechanism; readiness of the medical, materials, or industrial applications named in the press materials.

Main limitations: No peer review yet; no metabolic independence (ribosomes and enzymes are supplied); the headline cycle uses mechanical division; gene-driven division is lower-yield and needs added components; imperfect genome inheritance.

How much confidence should a general reader have? Reasonably high in the central engineering result: the authors ran the operational steps of a cell cycle together inside a defined synthetic droplet, though the work still awaits peer review. Medium-to-low confidence in the precise generation counts, division yields, and inheritance rates until peer review and independent replication. The paper does not claim that the system is living, self-sufficient, or self-evolving; those are scope boundaries, not low-confidence findings. Appropriate stance: an impressive step in building cells from known parts, not the creation of life.

Post-publication updates

- **Clarification · 2026-07-23**

Clarified the confidence framing: living, self-sufficient, and self-evolving cells are outside the paper's claims, not low-confidence findings. The assessment of the central engineering result is unchanged.

Sources

Gaut, Deich, Cash, Hoog, Engelhart & Adamala — 'A Chemically Defined Synthetic Cell Capable Of Growth And Replication', author-hosted preprint (not peer-reviewed), 2026

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